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Imaging and Therapy of Breast Cancer

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Las Cruces, New Mexico 88003

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13. ABSTRACT (Maximum 200 Words)

Several novel rhenium-estradiol complexes were synthesized as potential radiopharmaceuticals for the imaging and therapy of breast cancer. Novel palladium catalyzed carbon-carbon and carbon-nitrogen bond-forming chemistry was developed to synthesize phenyl and pyridyl hydrazine derivatives. Polymer-supported hydrazine derivatives were effective labeling substrates for synthesizing organoimidorhenium estradiol complexes. The estradiol-rhenium complexes were fully characterized structurally. The solubility and stability of the organoimidorhenium complexes in aqueous systems was investigated. Pyridylhydrazine derivatives of estradiol were found to form stable rhenium(I) carbonyl complexes, and are excellent candidates for continued investigation as diagnostic and therapeutic radiopharmaceuticals for breast cancer.

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INTRODUCTION:

This final report describes work completed on award number DAMD17-97-1-7120 during the time period May 12, 1997- May 12, 2001. This project resulted in the synthesis of several novel derivatives of estradiol that form complexes with rhenium, as potential radiopharmaceuticals for the imaging and therapy of breast cancer. Novel palladium catalyzed carbon-carbon and carbonnitrogen bond-forming chemistry was developed to synthesize a series of phenyl and pyridyl hydrazine derivatives. Polymer-supported hydrazine derivatives were found to be effective labeling substrates for the synthesis of organoimidorhenium estradiol complexes, and could be used as a kit formulation. The estradiol-rhenium complexes were fully characterized structurally, and submitted for receptor binding assays. The solubility and stability of the organoimidorhenium complexes in aqueous systems and in the presence of oxidants and amines was investigated. Pyridylhydrazine derivatives of estradiol were found to form extremely stable rhenium(I) carbonyl complexes, and are excellent candidates for continued investigation as diagnostic and therapeutic radiopharmaceuticals for breast cancer.

BODY:

The project was focused towards the accomplishment of these three technical objectives:

Technical Objective 1.

Synthesis of a series of polymer supported estradiol ligand precursors.

Technical Objective 2

Investigate and optimize the Re-imido forming reactions.

Technical Objective 3

Investigate physical characteristics and potential reactivity of Re-estradiol complexes.

variety of estradiol precursors were synthesized from $17-\alpha$ -ethynylestradiol and $17-\alpha$ ethenylestradiol by palladium-catalyzed carboncarbon bond forming reactions with the alkyne or alkene group respectively. The Sonogashira conditions (Pd(OAc)₂ CuI, Et₂NH) were effective for synthesizing aryl amine and hydrazine derivatives, and a manuscript describing these results was published.³ This paper presented the first examples of coupling an aryl halide substrate containing a free hydrazine group. The estradiol hydrazines were attached to commercially available carboxyl functionalized polymers by the formation of acyl hydrazine linkages. The Tentagel polymers were very effective, and exhibited solvent swelling in a variety of solvents. The polymer-supported acyl hydrazines were found to be effective for the synthesis of organoimidorhenium complexes. reactivity of these polymer-supported estradiol derivatives was investigated with various rhenium complexes, and the efficiency and rates of product formation were quantified for a series composed of different organoimido precursors with varying

polymeric backbones using the stable isotope of rhenium. The optimal reaction conditions for Relabeling were determined by monitoring the product formation using UV-Vis spectroscopy. Preparative scale reactions were performed and the product complexes were isolated in sufficient quantities for complete characterization. It was found that perrhenate (ReO4 the actual starting material for radiolabeling) with 188Re, could be used as the starting material by including triphenylphosphine hydrochloride and tetrabutylammonium chloride in the reaction mixture. This result validated the major theme of the proposal, that a polymersupported organoimido substrate could be used in a kit formulation to prepare Re-labeled estradiol without requiring separation of derivatives unlabeled estradiol. A manuscript describing these results was published.²

Further work involving the extrapolation of our results using the stable rhenium isotope to the ¹⁸⁸Re-radiolabeling of the polymer-supported estradiol hydrazine substrates are currently in progress in the laboratories of our collaboraters at Los Alamos National Laboratory. We anticipate a continuing relationship to further develop this technology.

We have structurally characterized a variety of rhenium complexes, 2,4,5 and investigated some reactions of the imido complexes that are possible in vivo. The hydrolysis of the organoimido linkage to the corresponding rhenium oxo complexes was a major concern, since this would cleave the estradiol group. We found that the hydrolysis of the imido group in the bis-triphenylphosphine complexes is generally slow, requiring several hours. However, unexpectedly, we found that N-hydroxy groups dramatically accelerate the hydrolysis via an oxidative pathway involving the

phosphine ligand followed by hydrolysis. Ligand substitution reactions were carried out to prepare dithiocarbamate complexes. These compounds were stable in the presence of mild oxidizing agents, but unfortunately underwent rapid hydrolysis in the presence of amine bases.

Recent work by Alberto and Schlibli has shown the excellent stability of rhenium(I) tricarbonyl complexes with pyridylhydrazines in water.8 In the last year of the project we focused on synthesizing pyridyl hydrazine derivatives of estradiol that would be capable of forming stable complexes. effort required the development of new methods for the amination of pyridyl substrates with hydrazine ^tBoc-protected pyridylhydrazine derivatives. derivatives were prepared from 2-pyridyl chlorides, bromides, and triflates using a one-step palladiumcatalyzed amination reaction with chelating phosphine ligands. The reaction conditions were optimized and the reactivity of a series of pyridyl substrates was investigated using an Argonaut Technology's Quest 210 parallel synthesizer. tert-butyl hydrazodiformate was found to be an excellent hydrazine substrate, and the resulting products were deprotected under mild conditions. This work was recently published in Organic Letters. 1

The catalytic amination chemistry of pyridyl halides was used to synthesize estradiol derivatives. The Sonogashira coupling reaction of 2-chloro-5-iodopyridine underwent selective C-C bond formation by substitution of the iodide with ethynylestradiol to give the 2-chloropyridyl derivative. The analogous Heck coupling of vinylestradiol produced the corresponding transalkene in 50% yield.

Our attempts to conduct the catalytic amination of the 2-chloropyridyl derivative were impeded by the unexpected fragmentation of the C-C bond to produce estrone as a major competing reaction. Control reactions demonstrated that palladium was essential for the degradation. The analogous alkene successfully underwent the amination in 66% yield, which indicates the unique sensitivity of the propargylic functional group under the elevated temperatures required for this reaction.

As an alternative route to the desired propargyl system we directly coupled ethynylestradiol with the 5-bromopyridyl hydrazine derivative using a Sonogashira reaction, and were delighted with the successful coupling yield of

this reaction (85%). Using vinylestradiol in an attempted Heck coupling with this substrate was unsuccessful, due to decomposition of the 5-bromopyridyl hydrazine at the elevated temperatures required for this reaction (100°C).

The estradiol pyridyl hydrazine derivatives were found to form stable complexes with the rhenium(I) tricarbonyl cation in water. The deprotection of the carbamate groups was facilitated by acidification with HCl. The complexes were stable to silica gel chromatography. A manuscript describing these results is currently in preparation. Samples will be further evaluation to determine their receptor binding affinity.

KEY RESEARCH ACCOMPLISHMENTS:

- New chemical methodology was developed, enabling for the first time the palladium catalyzed synthesis of aryl and pyridyl hydrazines.
- New technology for labeling estradiol with rhenium, compatible with instant kit type formulation, was developed.
- A variety of new rhenium-labeled estradiol derivatives were synthesized and characterized.
- The stability of organoimidorhenium linkages in aqueous systems was evaluated for the first time.
- A new class of estradiol derivatives possessing pyridyl hydrazine were synthesized, and have shown excellent stability, and indicate promise for continued evaluation as potential radiopharmaceuticals for diagnostic and therapeutic applications against breast cancer.

REPORTABLE OUTCOMES:

- Papers Published
- 1. Jeffrey B. Arterburn, Kalla Venkateswara Rao, Ranjit Ramdas, Benjamin R. Dible "Catalytic Amination of 2-Substituted Pyridines with Hydrazine Derivatives Organic Letters 2001, 3, 1351-1354.
- 2. Jeffrey B. Arterburn, Kalla Venkateswara Rao, and Marc C. Perry, Angewandte Chemie 2000, 39, 771-772. "Solid-Supported Hydrazine Substrate for Labeling Estradiol Ligands with Rhenium."
- 3. Jeffrey B. Arterburn, Kalla Venkateswara Rao, and Marc C. Perry, *Tetrahedron Letters* **2000**, 41, 839-842. "Novel 17α-Ethynylestradiol Derivatives: Sonogashira Couplings Using Unprotected Phenylhydrazines."
- 4. Jeffrey B. Arterburn, Kalla Venkateswara Rao,
 Donna M. Goreham, Marcela V. Valenzuela, Mylena
 S. Holguin, Keith A. Hall, Kevin C. Ott, and
 Jeffrey C. Bryan, Organometallics, 2000, 19,
 1789-1795. "Functionalized Re(V) Organoimido
 Complexes as Potential Radiopharmaceuticals
 (II). Synthesis, Structural Characterization,
 and Reactivity of N-Succinimidyl Ester
 Derivatives with Amines."
- 5. Jeffrey C. Bryan, Marc C. Perry, and Jeffrey B. Arterburn, Acta Crystallogr. (C), 1998, 54, 1607-1608.
 "Trichlorooxo(triphenylphosphine)(triphenylphosphine oxide)rhenium(V)."
- 6. Jeffrey B. Arterburn, Kalla Venkateswara Rao, Ranjit Ramdas "Synthesis of 2-pyridylhydrazine derivatives of estradiol as ligands for

- complexation with $\text{Re}(\text{CO})_3(\text{H}_2\text{O})_3^{+\prime\prime}$ Manuscript in preparation.
- 7. M. Fassbender , D.R. Phillips, K.C. Ott , and J.B. Arterburn "Non-carrier-added 186,188 Re labeled 17α -ethynylestradiol: a potential breast cancer therapy and imaging agent" J. Labeled Comp. Radiopharm. 2001 Manuscript in preparation.
- Degrees Awarded
- 1. Ph. D. Chemistry
 Marc C. Perry, Spring 2000
 Dissertation Title: "Rhenium-Catalyzed
 Heteroatom Transfer Reactions"
 Currently a postdoctoral associate at Texas A &
 M University.
- 2. M.S. Chemistry
 Ranjit Ramdas , Spring 2001
 Thesis Title: "The Catalytic Amination of
 Pyridyl Electrophiles with Protected
 Hydrazines; Synthesis of Pyridyl Derivatives of
 Estradiol" Currently employed.
- 3. B.S. Chemistry
 Marcela Valenzuela, Spring 2000. Currently a
 Ph.D. graduate student in chemistry at the
 University of Arizona.
- 4. B.S. Biochemistry, B.A. Chemistry
 Benjamin Dible, Spring 2001. Currently a Ph.D.
 graduate student in chemistry at the University
 of Utah.

- 5. B.S. Chemistry
 Reymundo Villa, Spring 2001. Currently a Ph.D.
 graduate student in chemistry at the University
 of California, Los Angeles.
- 6. B.S. Chemistry
 Cesear Corona, Spring 1999. Currently a Ph.D.
 graduate student in chemistry at New Mexico
 State University.
- 7. B.S. Chemical Engineering
 Mylena Holguin, Spring 1999. Currently employed.
- Other Funds Obtained

The successful results from this project were used as part of the Preliminary Results section of a new proposal to the National Institutes of Health titled "Development of New Methods for Radiolabeling with Tc and Re." which was funded for the period 3/1/00 to 2/28/04.

• Presentations

- 1. "Catalytic amination of 2-pyridyl systems with hydrazine derivatives" R. Ramdas, R. V. Kalla, B. R. Dible, J. B. Arterburn 221st American Chemical Society National Meeting, San Diego, CA April 1-5, 2001.
- 2. "Instant Kits for Labeling Estradiol with Rhenium." Jeffrey B. Arterburn, Era of Hope, Department of Defense (DOD) Breast Cancer Research Program (BCRP) Meeting. June 8-12, 2000. Atlanta, GA.
- 3. "Palladium-Catalyzed Synthesis of $17-\alpha-4-$ Hydrazidophenyl Estradiol Derivatives and Their Conversion to Organoimidorhenium(V) Complexes." Kalla Venkateswara Rao, Jeffrey B. Arterburn*, Marc C. Perry, Regional Meeting of the American Chemical Society, El Paso Texas, October 21-22 1999.
- 4. "Palladium-Catalyzed Synthesis of $17-\alpha-4-$ Hydrazidophenyl Estradiol Derivatives and Their Conversion to Organoimidorhenium(V) Complexes." Kalla Venkateswara Rao, Jeffrey B. Arterburn*, Marc C. Perry, 218th National Meeting of the American Chemical Society, New Orleans, LA August 22-26 1999.
- 5. "Solid-Supported "Instant Kits" for Labeling Estradiol Ligands with Rhenium."

 Jeffrey B. Arterburn, Kalla Venkateswara Rao, Marc C. Perry. 217th National Meeting of the American Chemical Society, Anaheim, CA March 21-25 1999.
- 6. "Rhenium-Catalyzed Reactions for Organic Synthesis" J. B. Arterburn, M.C. Perry,

- CU/Roche Colorado Symposium in Synthetic Chemistry, May 20-22, 1998.
- 7. "Non-carrier-added ^{186,188}Re labeled 17αethynylestradiol: a potential breast cancer therapy and imaging agent" M. Fassbender , D.R. Phillips, K.C. Ott , and J.B. Arterburn, 14th International Symposium on Radiopharmaceutical Chemistry, June 10-15, 2001 Interlaken, Switzerland.

CONCLUSIONS:

Novel palladium catalyzed carbon-carbon and carbon-nitrogen bond-forming chemistry was developed to enable the synthesis of a series of phenyl and pyridyl hydrazine derivatives. methodology was used for the synthesis of several new derivatives of estradiol that form complexes with rhenium, as potential radiopharmaceuticals for the imaging and therapy of breast cancer. Polymersupported hydrazine derivatives were found to be effective labeling substrates for the synthesis of organoimidorhenium estradiol complexes, and could be used as a kit formulation. This technology uses the starting complex perrhenate, which is the species used for 188Re labeling for nuclear medicine The estradiol-rhenium complexes were applications. fully characterized structurally, and submitted for receptor binding assays. The solubility and stability of the organoimidorhenium complexes in aqueous systems and in the presence of oxidants and amines was investigated. Hydrolysis of organoimido complexes containing phosphine ligands was accelerated by mild oxidizing agents. With ancillary dithiocarbamate ligands, hydrolysis of organoimido linkage was observed in the presence of amines. Pyridylhydrazine derivatives of estradiol were found to form extremely stable rhenium(I) carbonyl complexes, which are excellent candidates

for continued investigation as potential diagnostic and therapeutic radiopharmaceuticals for breast cancer.

REFERENCES:

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derivatives of estradiol as ligands for complexation with $\text{Re}(\text{CO})_3(\text{H}_2\text{O})_3^{+\prime\prime}$ Manuscript in preparation.

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- 8. R. Alberto, R. Schibli, A. Egli, A.P. Schubiger, U. Abram, T.A. Kaden *J. Am. Chem. Soc.* **1998**, *120*, 7987-7988. "A novel organometallic aqua complex of technetium for the labeling of biomolecules: Synthesis of [Tc-99m(OH₂)₃(CO)₃] from [(TcO₄)-Tc-99m] in aqueous solution and its reaction with a bifunctional ligand"

APPENDICES:

Papers 1-5

Catalytic Amination of 2-Substituted Pyridines with Hydrazine Derivatives

Jeffrey B. Arterburn, Kalla Venkateswara Rao, Ranjit Ramdas, and Benjamin R. Dible

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Catalytic Amination of 2-Substituted Pyridines with Hydrazine Derivatives

ORGANIC LETTERS

2001 Vol. 3, No. 9 1351–1354

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ABSTRACT

Protected pyridylhydrazine derivatives were prepared in a one-step palladium-catalyzed amination reaction using chelating phosphine ligands. 2-Pyridyl chlorides, bromides, and triflates were effective electrophiles in these reactions. Di-tert-butyl hydrazodiformate was an excellent hydrazine substrate, and the resulting products were deprotected under mild conditions. Catalytic amination provides a direct route to protected bifunctional hydrazinopyridine linkers that are suitable for metal-bioconjugate syntheses.

Hydrazinopyridines are important compounds for the synthesis of triazines and substituted pyridines as agrochemicals and pharmaceuticals.¹⁻⁴ 2-Pyridylhydrazines are particularly interesting synthetic targets as a result of their efficiency as ligands for a variety of metal complexes and potential applications in nuclear medicine.⁵⁻⁹ Direct displacement of 2-chloropyridines with hydrazine hydrate has been used for the synthesis of 2-pyridylhydrazine derivatives, and typical reaction conditions involve heating in highly corrosive, concentrated 85% hydrazine hydrate.^{10,11} These conditions

also generate the powerful reducing agent diimide, which rapidly reduces alkenes and alkynes.¹² Our interest in rhenium- and technetium-labeled biomolecules as potential radiopharmaceuticals^{13–15} led us to investigate metal-catalyzed amination as a possible alternative method for preparing protected 2-pyridylhydrazine derivatives that would be compatible with multistep syntheses. Catalytic amination of aryl halides and sulfonates has become an important method for preparing anilines and other arylamines.^{16–23}

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Chelating phosphine ligands enable Pd-catalyzed C-N bond formation of bromopyridines with alkyl and arylamines,²⁴ and sterically hindered monodentate biphenylphosphine ligands are useful for coupling chloropyridines.²⁰ Other examples of halopyridyl aminations catalyzed by Pd²⁵⁻³¹ and Ni^{32–37} have been reported. There are relatively few examples of catalytic amination reactions with hydrazine derivatives. The palladium-catalyzed amination of aryl halides with benzophenone hydrazone 1 has recently been reported. 20,38-40 The protected hydrazine tert-butylcarbazate 2 has also been used as a substrate for Pd-catalyzed aminations.41 Herein, we present the results of a comprehensive investigation of the catalytic amination of 2-pyridyl chlorides, bromides, and triflates with hydrazine substrates, 1-3, and provide a convenient general methodology for the synthesis of protected 2-hydrazinopyridine derivatives.

$$H_2N$$
 N H_2N H_2 H_2N H_3N $O-C(CH_3)_3$ H_3N $O-C(CH_3)_3$ H_3N $O-C(CH_3)_3$ H_3N $O-C(CH_3)_3$

The standard conditions^{38–40} for the N-arylation of 1 using 2 mol % Pd(OAc)₂, 2 mol % of the chelating ligand 2,2′-bis(diphenylphosphandanyl)-1,1′-binaphthyl (BINAP), and sodium *tert*-butoxide base in toluene were used for the pyridyl substrates **4a**–**f** as shown in Table 1. The hydrazone 1 was used as the limiting reagent to facilitate chromatographic purification of the products **5a**–**d**. 2-Chloropyridine **4a** was an effective substrate under these conditions. The reaction with 2-bromopyridine **4b** was faster and gave

Table 1. Coupling of 2-Pyridyl Systems with 1^a

entry	R	X	$product^b$	time (h)	yield (%)
1	Н	Cl, 4a	5a	16	70
2	H	Br, 4b	5a	8	95
3	Br	Br, 4c	5b	6	80
4	$\mathrm{CO_{2}C_{6}Cl_{5}}$	Cl, 4d	5c	14	42
5	H	OTf, 4e	5a	16	62
6	$\mathrm{CO_2C_2H_5}$	OTf, 4f	5d	16	72

 a All reactions were run with 2 mol % of Pd(OAc)2, 2 mol % of BINAP, 0.8 equiv of 1, 1.4 equiv of Na O'Bu, and 1 equiv of pyridine derivative, at 100 °C with 2 mL of toluene for 1 mM substrate. b All products were characterized by $^1\mathrm{H}$ and $^{13}\mathrm{C}$ NMR and HRMS spectroscopy methods. c Yields of isolated product.

improved yields, paralleling the normal relative reactivity of aryl halides. Changing the chelating ligand from BINAP to 1,1'-bis(diphenylphosphanyl) ferrocene (DPPF) in these cases resulted in significantly lower yields. The yields were also lower when Pd₂(dba)₃ was used as catalyst precursor. No reaction occurred using the catalytic system Ni(COD)₂/DPPF/NaO'Bu.³² Selective amination of the 2-position was observed for the Pd/BINAP-catalyzed reaction of 2,5-dibromopyridine **4c** with **1** in very good yield.

Activated ester derivatives of 6-hydrazidonicotinic acid are used as bifunctional labeling agents and are important synthetic compounds. 5-8 The Pd/BINAP-catalyzed amination of pentachlorophenyl ester analogue 4d with 1 gave a moderate yield of the desired compound 5c. We were interested in the possible catalytic amination of pyridyl triflates, since these compounds can be easily prepared from the hydroxypyridines⁴² and aryl triflates have been shown to be effective substrates for Pd-catalyzed amination with anilines and alkylamines. 43-46 The direct coupling of pyridyl triflates with amines is known;47,48 however, we observed no direct reaction between 1 and 4e after heating a toluene solution at 100 °C for 24 h. The pyridyl triflate 4e was converted to the hydrazone 5a by the Pd/BINAP catalyst. The yield of 5a from the triflate 4e was lower than that of the halogenated substrates but comparable to the amination of aryl triflates catalyzed by Pd(OAc)2/BINAP.43 The 2pyridyl triflates were stable under the basic reaction conditions, no competing cleavage to hydroxypyridine was

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*observed, and excess starting material was recovered at the end of the reaction. The presence of an ethyl ester substituent in the 5-postion **4f** resulted in a mild activation and improved the yield of the hydrazone product **5d**. The proposed mechanistic model for N-arylation involves oxidative addition as the rate-determining step. ⁴⁹ Coordination of pyridine has been shown to inhibit the Pd(0)/P(o-tolyl)₃-catalyzed amination of aryl bromides but does not displace bisphosphines. ²⁴ The electron-withdrawing ester substituent decreases the ligating ability of the pyridine further to enhance the rate of oxidative addition and also is expected to facilitate the subsequent reductive elimination that produces the C-N bond.

Electron-deficient aryl bromides and 4-nitro-2-bromopyridine have been shown to undergo Pd/DPPF amination with the protected hydrazine derivative *tert*-butylcarbazate **2**.⁴¹ We observed that 2-bromopyridine was coupled with **2** using 8 mol % Pd₂(dba)₃, 12 mol % DPPF, and Cs₂CO₃ in toluene. The N-pyridyl products underwent decomposition during attempted purification by silica gel chromatography. 2-Hydrazinopyridine **6** was obtained in an overall yield of 45% from 2-bromopyridine **4b** by hydrolysis of the crude reaction mixture using 20% HCl/EtOH, followed by neutralization and extraction with ether. No reaction of **2** with 2-bromopyridine was observed using the catalytic system Pd-(OAc)₂/BINAP/NaO'Bu that was effective with the hydrazone **1**, and no reaction occurred using Ni catalysts with various chelating ligands.³²⁻³⁷

The limited success we observed for coupling pyridyl substrates with 2 led us to investigate the possibility of Pd-catalyzed amination using the symmetrical hydrazine derivative di-tert-butyl hydrazodiformate 3, Table 2.

Catalytic N-arylations with protected amines such as carbamates have generally been difficult, ^{46,50} and no examples of catalytic C-N bond formation using 3 have been reported. Using 2 mol % Pd₂(dba)₃/3 mol % DPPF/Cs₂CO₃ in toluene, the reaction of 2-chloropyridine was very slow (entry 1). Increasing the catalyst to 8% decreased the reaction time and improved the yield significantly (entry 2). The more reactive 2-bromopyridine gave good yields using 2-4 mol % Pd; however, the optimum results were obtained using 8 mol % of Pd (entry 5). 2,5-Dibromopyridine 4c underwent selective coupling at the 2-position to produce 7b in high yield. The Pd/BINAP catalyst used for the synthesis of hydrazones 5a-d was not successful with 3.

Table 2. Coupling of 2-Pyridyl Systems with 3^a

entry	R	X	Pd/DPPF (mol %)	time (h)	yield ^{b,c} (%)
1	Н	Cl, 4a	2/3	16	7a , 30
2	H	Cl, 4a	8/12	16	7a , 70
3	\mathbf{H}	Br, 4b	2/3	16	7a , 61
4	\mathbf{H}	Br, 4b	4/6	8	7a , 61
5	H	Br, 4b	8/12	. 8	7a , 85
6	Br	Br, 4c	2/3	10	7b , 73
7	Br	Br, 4c	8/12	6	7b , 85
8	$\mathrm{CO_2C_6Cl_5}$	Cl, 4d	8/12	14	7c , 45
9	H	OTf, 4e	8/12	16	7a, 0
10	$\mathrm{CO_2C_2H_5}$	OTf, 4f	8/12	16	7d , 57

 a All reactions were run with specified mol % Pd/DPPF, 0.8 equiv of 3, 1 equiv of Cs₂CO₃, and 1 equiv of pyridine derivative at 100 °C with 2 mL of toluene for 1 mM substrate. b All products were characterized by $^1\mathrm{H}$ and $^{13}\mathrm{C}$ NMR and HRMS spectroscopy methods. c Yields of isolated product.

The pentachlorophenyl ester derivative of 2-chloropyridine 4d was coupled with 3 by Pd/DPPF to produce 7c in moderate yield. The unactivated pyridyl triflate 4e did not react under these conditions. Efforts to couple this substrate using the catalyst ligand combination Pd(OAc)₂/(o-biphenyl)-PCy₂, which is effective for aryl triflates,²⁰ were also unsuccessful. The introduction of an ethyl ester improved the reactivity of the pyridyl triflate 4f, such that 7d was formed in moderate yield using Pd/DPPF.

Scheme 2. Hydrolysis of the 'BOC-Protected Hydrazine 7b

These results demonstrate that Pd-catalyzed aminations with 1 and 3 are efficient methods for the preparation of protected hydrazone and *tert*-butyl carbamate ('BOC) protected hydrazine derivatives. To determine the relative ease of deprotection, we treated 5b and 7b with 20% HCl/EtOH. Under these conditions, the 'BOC protecting groups of 7b were completely hydrolyzed to furnish hydrazine dihydrochloride salt 8 in 75% yield. In contrast, the hydrazone 5b was stable under these conditions.

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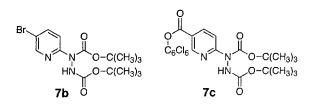


Figure 1. Protected bifunctional hydrazinopyridines.

These results illustrate the reactivity of 2-pyridyl electrophiles in Pd-catalyzed amination reactions and provide convenient methods for the synthesis of protected 2-pyridyl hydrazines. The reactions were successful using 2-pyridyl chlorides and triflates, although the best results were obtained with the bromides. The optimum reaction conditions for the pyridyl substrates with the hydrazine derivative 3 required a relatively high catalyst loading. The protected hydrazine derivative 3 was an excellent substrate for C-N bond formation and should be useful for the synthesis of a wide

variety of hydrazines via catalytic amination. The hydrolysis of the 'BOC protecting groups in 7a-d is considerably easier than for the hydrazone derivatives 5a-d. The catalytic amination of pyridyl electrophiles with 3 provides a direct route to protected bifunctional hydrazinopyridine linkers 7b and 7c, which are suitable for metal-bioconjugate syntheses. Efforts to further utilize these derivatives for the synthesis of radiopharmaceuticals are currently in progress.

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Supporting Information Available: Experimental procedures for **5a-d** and **7a-d** and analytical data for the compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

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Solid-Supported Hydrazine Substrate For Labeling Estradiol Ligands with Rhenium**

Jeffrey B. Arterburn,* Kalla Venkateswara Rao, and Marc C. Perry

Receptor-targeted radiopharmaceuticals offer great promise for the diagnostic imaging and therapy of tumors and other disease sites. Technetium-99m is readily available in nuclear medicine clinics throughout the world for diagnostic imaging applications, and the β -emitting radioisotopes of its congener, rhenium-186/188, are suitable for irradiating small to medium-sized tumors.[1] Radiolabeled bioligands such as steroids, peptides, and antibodies are capable of binding to receptors expressed by cancer cells, providing the selectivity needed for diagnostic and therapeutic applications.[2-4] The estrogen and progesterone steroid hormone receptors found in approximately two-thirds of breast tumors are suitable targets for steroid-based radiopharmaceuticals. [5] Radiopharmaceuticals with high specific activity are required, and the removal of all excess unlabeled ligand is essential to avoid competitive saturation of the binding sites of the ligand receptor. Herein we demonstrate a new strategy for labeling with rhenium using an organoimido-forming reaction of a polymer-supported hydrazine, which simultaneously establishes the steroidradioisotope linkage and releases the labeled steroid product into solution, thereby facilitating complete removal of all unlabeled ligand by simple filtration. The approach outlined here is uniquely amenable to the specific problem of developing "instant kits" for labeling low-capacity receptor ligands, and the technology is suitable for adaptation to a wide variety of different structural classes of ligands.

The 17α position of estradiol was selected as the site for appending the linking organoimido group, following the examples of organometallic steroid derivatives which exhibit high receptor binding affinities. We have previously synthesized highly functionalized organoimido complexes from substituted 1-acetyl 2-phenyldiazane (hydrazine derivatives) using carrier free trichlorooxobis(triphenylphosphane) rhenium(v), [ReOCl₃(PPh₃)₂]. Our approach required a convenient method for attaching pendant phenylhydrazine moieties to ethynylestradiol (1). The desired hydrazine 3 was obtained directly using a palladium-catalyzed coupling of ethynylestradiol (1) with 4-iodophenyl hydrazine (2) in diethylamine at ambient temperature in 87% yield (Scheme 1). The free hydrazine 3 was attached to Tentagel carboxy resin (loading capacity = 0.26 mmol g⁻¹) using (1-ben-

Scheme 1. a) 5 mol % Pd(OAc)₂, 10 mol % CuI, PPh₃, NHEt₂, 25 °C, 3 h, 87%; b) Tentagel carboxy resin, PyBOP, NEtiPr₂, CH₂Cl₂, 25 °C, 20 h, 100 % (based on loading capacity = 0.26 mmol g⁻¹); c) [ReOCl₃(PPh₃)₂], PPh₃, CH₂Cl₂, 40 °C, 3 h, 82 %.

zotriazolyl)oxy tris(pyrrolidino) phosphonium hexafluorophosphate (PyBOP) and diisopropylethylamine in dichloromethane to give the corresponding solid-supported acetyl hydrazine derivative 4. This reaction was monitored using FT-IR spectroscopy to follow the change in the carbonyl stretch from the free carboxylic acid (1737 cm⁻¹) to the carbohydrazide (1660 cm⁻¹), and the appearance of the characteristic aryl C-H bend at 1611 cm⁻¹ from the estradiol.

The organoimido-forming labeling reaction of solid-supported acetyl hydrazine 4 with [ReOCl₃(PPh₃)₂] (2.86 mm in CH₂Cl₂) and triphenylphosphane occurred readily in solution to produce the air- and moisture-stable complex 5 as an olive-colored solid in 82 % yield (Scheme 1). The product exhibited a single ³¹P NMR signal due to the coordinated triphenylphosphane ligands at $\delta = -20.4$ and displayed a characteristic UV/Vis absorption spectrum with maximum at $\lambda = 370$ nm ($\varepsilon = 16200$, CH₂Cl₂).

The previous example demonstrates the efficient reactivity of the polymer-supported hydrazines. The specific requirements for radiolabeling with rhenium and technetium involve highly dilute conditions, therefore a series of labeling reactions were carried out using the polymer-supported hydrazine 4 and dilute solutions of $[ReOCl_3(PPh_3)_2]$ from 10^{-5} to 10^{-6} M (Table 1). The formation of the organoimido complex 5 was followed spectroscopically by observing the absorption maximum at $\lambda = 370$ nm. The half lives for the labeling reactions $(t_{1/2} = 2 \text{ h})$ were unchanged over a 100- to 1000-fold excess of the support 4 relative to rhenium concentration. The yields of the reaction were similar when a solution of $[ReOCl_3(PPh_3)_2]$ prepared in situ from potassium perrhenate was used (entry 3, Table 1). [10] The organoimido

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Table 1. Rhenium labeling using the solid-supported hydrazine 4.

Entry	Concentration [10 ⁻⁵ M]	Conditions ^[a]	Time [h]	Yield [%]
1	2	A	2	53
			5	72
2	0.2	В	2	50
			5	7 0
3	9.3	С	2	54
			5	7 0
4	2	D	2	55
			5	65

[a] Reactions carried out in CH₂Cl₂ (10 mL) at 40 °C. Reactant ratios used: A: $4/PPh_3/[ReOCl_3(PPh_3)_2] = 100/100/1$; B: $4/PPh_3/[ReOCl_3(PPh_3)_2] = 1000/1000/1$; C: $4/PPh_3/KReO_4 = 100/100/1$; D: $4/HPPh_3Cl/Bu_4NReO_4 = 100/100/1$.

complex 5 was also prepared using a one-pot procedure starting with tetrabutylammonium perrhenate and triphenyl-phosphane hydrochloride in dichloromethane (entry 4, Table 1).

These examples illustrate a new strategy for labeling estradiol ligands with rhenium using a solid-supported hydrazine substrate, and this chemistry should also be successful for preparing technetium analogs. [12] The ability to use perrhenate and pertechnetate salts for labeling is particularly advantageous, since these species are obtained directly from the radionuclide generators. The efficiency and convenience of this approach can be extrapolated to a new generation of rhenium and technetium complexes for diagnostic and therapeutic applications in nuclear medicine. Further studies that are currently in progress involve evaluation of the receptor binding affinity and in vivo stability of these estradiol derivatives, and the extension of this technology to other low-capacity receptor systems.

Experimental Section

- 4: To a suspension of Tentagel Carboxy resin (1.0 g, 0.26 mmol) in dichloromethane (50 mL) was added PyBOP (405 mg, 0.78 mmol) followed by hydrazine 3 and diisopropylethylamine (0.3 mL). The resulting suspension was stirred at 25 °C for 20 h. The reaction product was filtered to give the yellow polymer support 4. FT-IR (KBr): $\bar{v} = 3443$, 2869, 1652, 1611, 1104 cm⁻¹.
- 5: A suspension of 4 (110 mg, 0.0286 mmol), triphenylphosphane (7.5 mg, 0.0286 mmol), and [ReOCl₃(PPh₃)₂] (23.8 mg, 0.0286 mmol) in dichloromethane (10 mL) was heated at 40 °C for 3 h. The reaction mixture was filtered, and washed thoroughly with dichloromethane. The combined organic layers were concentrated, and the product was precipitated from dichloromethane/hexanes and recrystallized to provide the complex 5 (28 mg, 82%) as olive-green crystals containing CH₂Cl₂. Elemental analysis for C₆₂H₅₇Cl₃NO₂P₂Re·0.5 CH₂Cl₂: calcd: C 60.23, H 4.61, N 1.12; found: C 60.29, H 4.66, N 1.50. Spectral data is provided in the Supporting Information.

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Regioselective Lactonization of Tetrasialic Acid**

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Polysialic acids (PSAs) are polymers of N-acetylneuraminic acid. Depending on their glycosidic linkages, these sugar polymers exist in nature as α -2,8-, α -2,9-, and α -2,8/2,9-linked polysaccharides. PSAs have been reported to demonstrate many important biological functions. For example, α -2,8-PSA is mainly linked to the neural cell adhesion molecule (N-CAM). This homopolymer of sialic acid has been implicated in reducing N-CAM adhesion; removal of the PSA increases the adhesive capability of N-CAM. In addition, α -2,8- and α -2,9-PSAs are the capsular polysaccharides of, respectively, serogroups B and C of *Neisseria meningitidis*, a leading worldwide cause of meningitis and rapidly fatal sepsis in otherwise healthy individuals. In the sugar of the polysaccharides of the polysacch

Structural diversities of PSAs are even more complicated with the possibility of PSA lactonizations. For α -2,8-PSA, the C-2 carboxylic acid of one residue can condense with the C-9 hydroxyl group of an adjacent residue to generate a δ -lactone under acidic conditions. Such δ -lactonizations have also been

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Novel 17α-ethynylestradiol derivatives: Sonogashira couplings using unprotected phenylhydrazines

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Abstract

The Pd/Cu catalyzed coupling of 17α -ethynylestradiol with halogenated amino-substrates was investigated. Iodophenylhydrazine and its protected derivatives reacted with 17α -ethynylestradiol to give 4-hydrazinophenyl derivatives without any degradation of the hydrazine group. Unprotected 3-, and 4-iodoaniline reacted similarly to produce the aminophenyl-derivatives. Protection of the amino group of halogenated benzylamines was required for alkyne coupling reactions, in order to avoid competing *ortho*-palladation of the benzylamine substrates. © 2000 Elsevier Science Ltd. All rights reserved.

Keywords: amine; hydrazine; palladium; steroid.

There has been a great deal of interest in the synthesis of estradiol derivatives with enhanced binding affinity for the estrogen receptor. We desired a direct route for preparing aniline, benzylamine, and phenylhydrazine derivatives appended to the ethynyl group of 17α -ethynylestradiol 1a. The Sonogashira coupling of alkynes with aryl iodides and bromides catalyzed by Pd(O) and Cu(I) provides a mild, and efficient way to synthesize aryl alkynes; 1-5 however, very few examples of the coupling reaction with amino-substrates have been reported, 6-8 and there are no examples of the alkyne coupling reaction with halogenated unprotected phenylhydrazines. Herein we describe the synthesis of eight derivatives of 17α -ethynylestradiol, and identify the scope and limitations of the Sonogashira coupling reaction with hydrazine, and amine-containing substrates.

The Pd/Cu(I) catalyzed coupling reaction between **1a** and 3-iodoaniline **2a** in diethylamine at 25°C gave the desired *m*-aniline derivative **3a** in excellent yield (Scheme 1). The coupling was also successful using 4-iodoaniline **2b** to give the *p*-substituted compound **3b** in 89% yield (Table 1). Attempts to couple **1a** with 3-iodobenzylamine or 4-bromobenzylamine directly using these conditions were unsuccessful, and only self-coupling of the alkyne and degradation of the starting benzylic amines were observed. Benzylamines have been reported to undergo a direct *ortho*-palladation reaction with Pd(OAc)₂, ^{9,10}

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which would prevent the desired catalytic cycle for alkyne coupling. The first step in the *ortho*-palladation involves coordination of the benzylamine, therefore we converted the amine to a non-coordinating derivative. The 'butoxycarbonyl-protected (BOC-) 3-iodobenzylamine 2d reacted smoothly with 1a to produce the coupled product 3d in 75% yield. Deprotection of this compound in 3N HCl/EtOAc gave the benzylic amine derivative 3c in 80% yield. The coupling reaction was also successful using the BOC-protected 4-bromobenzylamine 2e, but due to the reduced reactivity of aryl bromides, heating for 4 h at 60°C was required for complete conversion.

$$\begin{array}{c} \text{OH} \\ \text{C=C-H} \\ \hline \\ \text{Smol}\% \ \text{Pd}(\text{OAc})_2 \\ \text{PPh}_3, \ 10 \text{mol}\% \ \text{Cul} \\ (C_2H_5)_2\text{NH}, \ 3h, \ rt \\ \\ \text{1b: R = TBDMS} \\ \end{array}$$

Scheme 1.

The catalytic coupling of hydrazine derivatives is complicated by the competing possibilities of hydrogen transfer chemistry, and reductive cleavage of the N–N bond. The monoarylhydrazine 2f underwent the coupling reaction catalyzed by Pd/Cu(I) to give the *p*-phenylhydrazine derivative 3f in 87% yield. This is the first example of a coupling reaction involving an unprotected mono-substituted hydrazine. The acetyl- and BOC-protected hydrazine derivatives 2g, and 2h were also converted to the alkyne products in very good yields. The 'butyldimethylsilyl ether group (TBDMS) of the protected estradiol derivative 1b and the BOC group of 2h were stable under the mild reaction conditions which were used to produce 3h. The hydrazine products were easily isolated by evaporation of the diethylamine solvent followed by silica gel chromatography eluted with 5% CH₃OH/CH₂Cl₂.

In summary, the Sonogashira coupling with amine and hydrazine substrates provides an efficient method for derivatizing 17α -ethynylestradiol. The mild reaction conditions, protecting group tolerance, simple workup procedures, and very good product yields are particularly advantageous. No complications from side reactions of the hydrazine group, or degradation of the sensitive phenol and tertiary alcohol functional groups of the steroid were observed. Benzylic amine substrates must first be protected before undergoing the coupling reaction, and the BOC group can be easily removed from the alkyne product. This procedure should be of general utility for the synthesis of conjugated aniline, benzylamine, and phenylhydrazine derivatives.

Typical experimental procedure: Synthesis of 17α-(3'-aminophenyl)ethynylestradiol (**3a**): A solution of palladium acetate (6.0 mg, 0.025 mmol) and triphenylphosphine (13 mg, 0.05 mmol) in diethylamine (3 mL) was stirred under argon for 10 min. Copper(I) iodide (10 mg, 0.05 mmol) and 3-iodoaniline **2a** (110 mg, 0.50 mmol) were added; after 5 min **1a** (148 mg, 0.50 mmol) was added and the reaction stirred for 3 h. Diethylamine was removed in vacuo. The residue was chromatographed on a silica gel column (25 g), eluted with 5% CH₃OH/CH₂Cl₂ to give **3a** (170 mg, 89% yield) as a white solid: mp 151–153°C; FT-IR (KBr, cm⁻¹) 3376, 1601, 1499, 786, 688; ¹H NMR (25% DMSO- d_6 /CDCl₃) δ 8.62 (br s, 1H), 7.08 (d, J=8.8 Hz, 2H), 7.02 (t, J₁=9.0 Hz, J₂=8.2 Hz, 1H), 6.80–6.45 (m, 5H), 4.95 (br s, 1H), 2.76 (s, 2H), 2.40–1.20 (m, 14H), 0.88 (s, 3H); ¹³C NMR δ 154.39, 146.87, 137.09, 130.41, 128.50, 125.61, 123.28, 120.06, 116.74, 114.67, 114.17, 112.35, 92.95, 84.68, 78.68, 49.05, 46.96, 43.15, 39.9, 39.5, 32.54, 29.05, 26.75, 25.99, 22.33, 12.44; anal. calcd for C₂₆H₂₉NO₂–0.5H₂O: C, 78.75; H, 7.63; N, 3.53. Found: C, 78.47; H, 7.49; N, 3.81.

 $Table \ 1$ Coupling of $17\alpha\text{-ethynylestradiol} \ \textbf{1a}$ with amines, and hydrazines a

Entry	Aryl halide	Coupled product	Yield (%) ^b
1	ı—⟨	OH C≡C NH ₂	90
2	I—√NH ₂ 2b	OH C≡C NH ₂	89
3	ı————NH₂	OH C=C-NH ₂	0
4	I———NHBOC	OH C=C-NHBOC	75
5	Br—NHBOC	OH C=C-NHBOC	76
6	I——N−NH ₂ 2f	OH C≡C N-NH ₂	87
7	I——N-N-C-CH ₃	OH C≡C — N-N-C-CH ₃ 3g	76
8	I——N-NHBOC	OH C≡C——N-NHBOC H-NHBOC	80°

^a All reactions were carried out under argon at room temperature except entry 5 which was heated to 60 °C. ^b All yields are of pure products isolated by silica gel column chromatography, and all the products gave satisfactory spectral and analytical data. ^c Alkyne **1b** was used

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Functionalized Rhenium(V) Organoimido Complexes as Potential Radiopharmaceuticals. 2. Synthesis, Structural Characterization, and Reactivity of N-Succinimidyl Ester Derivatives with Amines

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Organoimidorhenium(V) complexes were synthesized as potential labeling agents for biologically relevant organic amines using the preconjugate approach. The bistriphenylphosphine organoimidorhenium N-succinimidyl ester complex $Cl_3(PPh_3)_2Re=N-C_6H_4CO_2N-(COCH_2)_2$ (2) was synthesized and characterized by single-crystal X-ray analysis. Complex 2 was coupled in aqueous dimethylformamide solvent with a series of primary and secondary amines, amino acids, and a biotin—ethylenediamine derivative to give the corresponding amide complexes in good yields. These results demonstrate that the organoimido linkage is resistant toward hydrolysis and stable in the presence of more basic alkylamines. An unusual oxygen atom transfer reaction was observed between the byproduct N-hydroxysuccinimide and triphenylphosphine ligands when dichloromethane was used as solvent. The dithiocarbamate complexes $Cl[Et_2NCS_2]_2Re=N-C_6H_4CO_2N(COCH_2)_2$ (3) and $Ol(Et_2NCS_2)_2Re=N-C_6H_4-CO_2N(COCH_2)_2$ (4) were also synthesized from 2. These complexes were unaffected by N-hydroxysuccinimide, but were not suitable for labeling due to hydrolysis of the organoimido groups under the reaction conditions.

Introduction

Biomolecules labeled with organometallic and coordination complexes have been developed for use as electron-transfer mediators, as probes for the active sites of enzymes, and for protein structural resolution with X-ray diffraction and electron microscopy.1 The synthesis of radiopharmaceuticals represents another area where new technologies are necessary to overcome the challenges associated with labeling biomolecules with radioactive metals.2 Peptides, antibodies, and steroids are all potential substrates for labeling with radioactive metals such as Tc-99m and Re-186/188 for diagnostic imaging and targeted radiotherapy, respectively. The ideal radiopharmaceutical would be easy to synthesize, exhibit stability and selective biodistribution in vivo, and then clear the body after the clinical procedure. These stringent requirements can ultimately be attained through multifaceted approaches leading to a diverse variety of candidate complexes for clinical trials.

Three general methods for producing metal bioconjugates are available: (1) direct labeling of biomolecules that contain a natural ligating or reactive group such as thiols; (2) the postconjugate approach where the biomolecule is first attached to a modified ligand that is then complexed with the metal; and (3) the preconjugate approach, where a metal complex is formed first and then attached to the biomolecule as an intact unit. With direct labeling it is difficult to control the site of attachment, the structure of the biomolecule can be disrupted, and redox changes at the metal can occur. The postconjugate approach provides a controlled ligand environment, but remains subject to problems during

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complex formation. The preconjugate approach offers greater control through the use of a purified metal complex, which only undergoes labeling with appropriately matched functional groups on the biomolecule. Amine side chain groups are commonly found on the biomolecules of interest and offer convenient sites for conjugation to a modified ligand system, due to the ease of formation of amide bonds with various activated carboxyl derivatives. Activated esters have been used to attach chelating ligands,3 metal complexes,4 and groups such as hydrazinopyridine to biomolecules.5 The N-succinimidyl ester group is a convenient activating group for the acylation of amines and has been employed in the preconjugate approach to incorporate a variety of different classes of metal complexes, 6 including rhenium and technetium.7

We have investigated the suitability of the multiple bonding imido ligand for Tc and Re and recently reported the synthesis of aryl organoimido Re(V) complexes containing remotely functionalized carboxylic acid groups 1.8 These compounds are stable to air as solids and in solution, and Re-188 complexes can be synthesized at tracer levels and purified by HPLC chromatography using aqueous acetonitrile eluent. The multiply bonded organoimido group provides a direct covalent linkage between the organic and metal components. With this mode of attachment it is possible to fine-tune desirable properties of size, charge, and lipophilicity by varying labile ligands in the coordination sphere, an option unavailable to chelates. Therefore we have considered the development of organoimido com-

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Scheme 1

$$\begin{array}{c} N_{3} & & \\$$

plexes that are remotely functionalized with active ester analogues which would be suitable for labeling amines by the preconjugate approach. This approach requires that the organoimido linkage be resistant to hydrolysis under aqueous conditions and exhibit stability in the presence of alkylamines, which are both stronger bases and potential ligands. We describe here the synthesis and X-ray stucture of the activated organoimido Nsuccinimidyl ester 2 and dithiocarbamate derivatives 3 and 4, and an investigation of their reactivity with amines, amino acids, and a biotin derivative targeted for monoclonal antibody-avidin conjugates.

Results and Discussion

Synthesis and Characterization of Organoimido Succinimidal Esters 2-4. Iminophosphoranes are known to react with rhenium(V) oxo complexes to form arylimidorhenium(V) complexes and triphenylphosphine oxide.9 The bis-triphenylphosphine organoimidorhenium N-succinimidyl ester complex Cl₃(PPh₃)₂- $Re=N-C_6H_4CO_2N(COCH_2)_2$ (2) was prepared in 91% yield by heating the rhenium(V) oxo complex ReOCl₃-(PPh₃)₂ in benzene for 1 h with the iminophosphorane derived from N-succinimidyl-4-azidobenzoate (Scheme 1).¹⁰ No precautions to dry the solvent or exclude air were required. The complex exhibits a single ³¹P{¹H} NMR signal at δ -23.7 for the equivalent *trans* triphenylphosphine ligands. The complex was fully characterized by ¹H and ¹³C NMR and elemental analysis. The FT-IR absorbances of the ester and imide carbonyl groups occur at 1770 and 1743 cm⁻¹, respectively. The complex exhibits a characteristic UV-vis absorbance at $\lambda = 336$ nm, $\epsilon = 14\,030$ in CH₂Cl₂ and is easily

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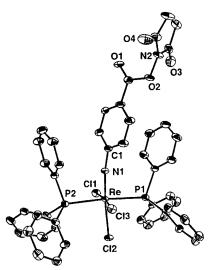


Figure 1. ORTEP drawing (50% probability ellipsoids) of $Cl_3(PPh_3)_2Re = N - C_6H_4CO_2N(COCH_2)_2 \cdot (CH_2Cl_2)_{1.43}$ (EtOH)_{0.57}. Hydrogen atoms and disordered lattice molecules are omitted.

Table 1. Summary of Crystallographic Data

I table it building of c	-J
chemical formula	$C_{49.57}H_{44.28}Cl_{5.86}N_2O_{4.57}P_2Re$
temp (K)	203
space group	$P\bar{1}$ (No. 2)
cryst dimens (mm)	$0.18 \times 0.23 \times 0.68$
a, A	12.703(3)
b, A	14.250(3)
c, Å	14.930(3)
α, deg	100.04(3)
β , deg	105.40(3)
γ, deg	101.23(3)
cryst syst	triclinic
Z, volume (Å ³)	2, 1197.0
density (calcd) (g/cm ³)	1.60
abs coeff (mm ⁻¹)	2.88
transmission factors	0.592, 0.719
θ range for data collection (deg)	2.0-25.0
limiting indices	$-15 \le h \le 0, -16 \le k \le 16,$
minimg marces	$-17 \le l \le 17$
no, of refins collected	8965
	8537, 0.029
no. of indepdt reflns, $R_{ m int}$	•
R^a	0.034

 $^{a}R = (\sum |[F_{\text{obs}} - F_{\text{calc}}]|)/(\sum F_{\text{obs}}).$

Table 2. Selected Bond Lengths (Å) and Angles (deg) for 2

bond di	istances	bond an	gles
Re-N1	1.721(4)	Re-N-C1	171.4(4)
Re-Cl1	2.440(1)	P1-Re-P2	175.3(1)
Re-Cl2	2.411(1)	Cl2-Re-Cl3	85.4(1)
Re-P1	2.524(2)	Cl1-Re-N	93.9(1)
Re-P2	2.518(2)	Cl1-Re-P1	89.7(1)
N1-C1	1.380(6)	Cl1-Re-P2	90.0(1)
O1-C7	1.211(6)	Cl1-Re-Cl2	90.4(1)

distinguished from the oxo complex ReOCl₃(PPh₃)₂, which absorbs at 272 nm.

The structure of the complex 2 (Figure 1) consists of an octahedral arrangement of mer-cis-oriented chloride ligands, trans-triphenylphosphine ligands, and the imido ligand. This ligand arrangement around the rhenium is similar to that of previously reported phenylimido complexes and is consistent with spectroscopic data for the complex in solution. A summary of crystallographic data is reported in Table 1. Selected bond lengths and angles are reported in Table 2. The Re-N bond length of 1.721(4) A and the nearly linear Re-N-

C1 organoimido bond angle of 171.4(4)° are typical of multiple bonded rhenium(V) monoimido complexes (1.69-1.75 Å, 156-180°).11 The Re-Cl and Re-P distances are similar to those reported for the closely related complex [Re(NPh)Cl₃(PPh₃)₂]. The N-succinimidyl ester carbonyl group of 2 is situated remotely from the coordination environment of the metal and is not perturbed by it.

The monomeric dithiocarbamate complex Cl[Et2- NCS_2 ₂ $Re=N-C_6H_4CO_2N(COCH_2)_2$ (3) was prepared in 84% yield by heating a solution of 2 with tetraethylthiuramdisulfide in acetone for 1 h (Scheme 1).13 The 1H NMR spectra of 3 in CDCl3 at 22 °C shows a single triplet for the four methyl groups and a multiplet for the inequivalent CH2 groups due to slow rotation about the C-N bond of the dithiocarbamate ligands. The ethyl groups in 3 give rise to two 13 C NMR peaks at δ 12.42 and 45.16. The FT-IR spectra show strong absorbances at 1531 and 997 $\rm cm^{-1}$ associated with the $\rm \bar{C}{-}N$ and $\rm C{-}S$ bonds of the dithiocarbamate ligand. The absorbances for the ester and imide carbonyls of 3 appeared at 1769 and 1741 cm^{-1} and are very close to the corresponding carbonyl groups of complex 2. Complex 3 exhibits a UVvis absorbance in CH₂Cl₂ at 418 nm, $\epsilon = 4710$ that is easily distinguished from the oxo complex Cl[Et2NCS2]2-Re=O, which absorbs at 344 nm.

The tetrakis(dithiocarbamate) μ -oxo-dirhenium complex $O[(Et_2NCS_2)_2Re=N-NC_6H_4CO_2N(COCH_2)_2]_2$ (4) was prepared in 73% yield by treating 3 with sodium carbonate in 0.1% v/v aqueous acetone and heating for 1 h (Scheme 1).13 The 1H NMR spectra of 4 in CDCl3 at 22 °C shows a single triplet for the four methyl groups and a multiplet for the CH2 groups of the dithiocarbamate ligand. The FT-IR spectra of 4 shows strong absorbances of the dithiocarbamate ligand at 1500 and 1072 cm⁻¹ and also an absorbance at 684 cm⁻¹ due to the bridging μ -oxo Re-O-Re group. The absorbance of the ester carbonyl in 4 is shifted to 1780 cm⁻¹, an increase of +10 cm⁻¹ compared with complexes 2 and 3. This shift indicates the apparently greater electron-withdrawing effect of the bridging oxo group in comparison with the chloride ligand, which influences the remote conjugated carbonyl through the phenylimido group. Complex 4 showed a UV-vis absorbance at 444 nm in CH₂Cl₂; however this absorbance exhibited a nonlinear dependence on concentration. Similar deviations from the Beer-Lambert law have been observed in the electronic spectra of related oxo-bridged organoimido molybdenum dimers and were attributed to an equilibrium disproportionation reaction.14 The corresponding oxo dimer O[(Et2NCS2)2Re=O]2 absorbs at 388 nm in CH₂Cl₂ and is easily distinguished from the organoimido complex 4.

Stability Studies of Organoimido Succinimidyl Esters 2-4. The characteristic UV-vis absorbances of the organoimido complexes were useful for monitoring the stability of the organoimido bond toward hydrolysis in solution. For the developmental stage of this project,

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the stability of the complexes over the course of a 3 h time period was taken as representative of the necessary time for preparing, administering, and imaging a potential organoimido radiopharmaceutical. Solutions of all three of the organoimido complexes in CH2Cl2 or CDCl3 were stable for several days at room temperature, exhibiting no change in either their absorbance spectra or their ¹H NMR. The complex 2 was not soluble in water alone, but dilute solutions (10^{-5} M) in 10% DMF/ H₂O were prepared by first dissolving the complex in DMF and then diluting with H2O. The complex 2 decomposed slowly in the aqueous media, exhibiting 30% hydrolysis after 3 h. The simple oxo complex ReOCl₃(PPh₃)₂ resulting from hydrolysis of the organoimido bond was observed by UV-vis, and corresponding amounts of the aniline H₂NC₆H₄CO₂N(COCH₂)₂ were isolated. Hydrolysis of the succinimide ester group was not observed during the 3 h time period. The monomeric chloro dithiocarbamate complex 3 rapidly dimerized in 10% DMF/H₂O to form 4. This reaction is similar to the preparative dimerization that is carried out in aqueous acetone in the presence of base and precludes the formation of monomeric complexes from 3 in the presence of H_2O .¹³ The μ -oxo dimer complex 4 was very stable in the aqueous DMF and exhibited no hydrolysis of the organoimido bond after 3 h, as evidenced by NMR and UV-vis.

Reactivity of N-Succinimidyl Ester 2 with Amines. The amine coupling reactions of 2 were initially conducted with isopropylamine using CH_2Cl_2 as solvent (eq 1). In this reaction the nucleophilic amine

undergoes acylation with the succinimidyl ester complex 2 to produce the amide product 5a and 1 equiv of N-hydroxysuccinimide HON(COCH₂)₂. Because the ultimate goal was to develop practical amine labeling agents, no efforts were taken to dry solvents and reagents or to exclude air from the reactions. The amide product 5a was isolated by precipitation with hexanes after concentrating the solvent. The complex 5a was characterized by ¹H and ¹³C NMR and elemental analysis. The FT-IR of 5a shows the absorbance of the amide C=O group at 1639 cm⁻¹. The isolated yields of product 5a from repeated experiments in CH₂Cl₂ were consistently below 30% and were accompanied by large amounts of triphenylphosphine oxide (OPPh₃), succinimide HN(COCH₂)₂, and the hydrolyzed amide H₂NC₆H₄CONHCH(CH₃)₂. The yields of product **5a** were not improved when this reaction was repeated using anhydrous CH₂Cl₂ under an atmosphere of argon. The isolated product 5a was found to be stable in the presence of excess isopropylamine in CH₂Cl₂ solution for at least 6 h.

A control experiment was performed to evaluate the stability of 5a in the presence of $HON(COCH_2)_2$, the reaction byproduct of amide formation. A 10^{-2} M solution of the amide complex 5a in CH_2Cl_2 was treated with 2 equiv of $HON(COCH_2)_2$, and the UV-vis spectrum was monitored during the course of the reaction. The

absorbance due to the coordinated PPh₃ ligands at 264 nm decreased in intensity and was completely replaced by the appearance of OPPh₃ after 1 h. The absorbance at 342 nm shifted slightly to 362 nm, suggesting the possible existence of a phosphine oxide complex with an intact organoimido bond. The addition of water eliminated the absorbance at 362 nm and caused decomposition of the complex along with hydrolysis of the organoimido group to produce the aniline derivative H₂NC₆H₄CONHCH(CH₃)₂, which was isolated from the reaction mixture in 68% yield after silica gel chromatography (eq 2). Attempts to isolate the putative phos-

phine oxide complex from these reactions and preparations carried out with exclusion of O2 and H2O were unsuccessful. The PPh3 ligands in complex 5a were not displaced by the addition of excess OPPh3. The low amide coupling yields of 5a with isopropylamine in CH₂Cl₂ can therefore be attributed to oxidation of the coordinated phosphine ligands by HON(COCH₂)₂, which is released after each coupling event, followed by hydrolysis of the imido bond, which occurs faster in complexes containing phosphine oxide ligands. The oxygen atom transfer chemistry to the coordinated PPh₃ from N-hydroxysuccinimide in CH2Cl2 was unexpected and to the best of our knowledge unprecedented in the literature, although various other N-oxides are recognized as thermodynamically favorable oxidants toward oxidation of PPh₃.15

The yield of the coupling reaction was significantly improved by changing the solvent to DMF, and complex 2 reacted with a series of amines to produce the corresponding amide complexes 5a-i, Table 3. In a typical reaction, complex 2 was treated with 10 equiv of the amine in DMF (0.025 M) at 25 °C for 10 min, followed by the addition of water to precipitate the amide products. Two factors contributed to the dramatic success of this procedure: the enhanced rate of amide formation in DMF and the ability to precipitate the products with H₂O, which concentrated the HON-(COCH₂)₂ in the aqueous phase, thereby preventing it from oxidizing bound PPh3 ligands of the metal products. The yields of the organoimido amide products from primary and secondary amines using this procedure varied from 67% to 78%. Complexes 5a, 5d, 5e, and 5f were recrystallized from CH2Cl2, and the solids retained some CH2Cl2 solvent, which was not removed by drying in vacuo and was evident in the NMR spectra and elemental analyses of these compounds. The ethyl ester hydrochloride salts of alanine and cysteine were coupled under similar conditions in the presence of added triethylamine, producing amides 5g,h in 67% and 74% yields, respectively. The success of the coupling reaction

Table 3. Coupling of 1° and 2° Amines with 2^{a}

entry	amine	product-amide	yield (%)
1	isopropylamine	5a : R' = H	70
	1 10	$R = -CH(CH_3)_2$	
2	2-amino-3-methylbutane	$\mathbf{5b} \colon \mathbf{R'} = \mathbf{H}$	69
		$R = -CH(CH_3)CH(CH_3)_2$	
3	phenethylamine	$\mathbf{5c}$: $\mathbf{R'} = \mathbf{H}$	67
		$R = -CH_2CH_2 - C_6H_5$	
4	benzylamine	$\mathbf{5d} \colon \mathbf{R'} = \mathbf{H}$	72
		$\mathrm{R} = -\mathrm{CH_2} - \mathrm{C_6H_5}$	
5	pyrrolidine	5e : $R = R' = N(CH_2CH_2)_2$	70
6	morpholine	5f : $R' = R = N(CH_2CH_2)_2O$	7 8
7	alanine ethyl ester hydrochloride ^b	$\mathbf{5g}: \mathbf{R'} = \mathbf{H}, \mathbf{R} = -\mathbf{CH}(\mathbf{CH}_3)\mathbf{CO}_2\mathbf{CH}_2\mathbf{CH}_3$	67
8	cysteine ethyl ester hydrochloride ^b	$\mathbf{5h}: R' = H, R = CH(CH_2SH)CO_2CH_2CH_3$	74
9	6^{b}	5i: R' = H	49
		$R = -(CH_2)_2NHC(O)$ biotin	

^a Reactions were carried out in DMF, and the product was precipitated with water. ^b Triethylamine (2 mmol) was added in the reaction

in the presence of the free sulfhydryl group of cysteine is particularly notable considering the strong potential for ligation of this group. Solutions of the cysteine complex **5h** did exhibit slow decomposition after several hours at ambient temperature, which presumably could be initiated by intermolecular ligand substitution reactions involving the sulfhydryl group.

There has been a great deal of interest in preparing labeled biotin derivatives for use in radioimmunodetection and radioimmunotherapy. ¹⁶ In the pretargeting approach, monoclonal antibody—avidin conjugates are allowed to localize at their target site, followed by treatment with a radiolabeled biotin derivitive, which is then selectively accumulated due to the high binding affinity of avidin for biotin. The ethylenediamine derivative of biotin 6 was prepared following the literature procedure. ¹⁷ The coupling reaction of 2 with 6 was performed in DMF as described above and afforded the biotin—amide derivative 5i in 49% isolated yield (eq 3). The amide coupling proceeded to completion; however

the lower isolated yield in this case can be attributed to the solubility characteristics of the biotin moiety in 5i, which adversely affected the precipitation of the complex. The complex was characterized by ³¹P{¹H}, ¹H, and ¹³C NMR.

Reactivity of Dithiocarbamate Complexes 3 and 4. Recognizing the problems associated with oxidation of the phosphine ligands by *N*-hydroxysuccinimide, we

decided to investigate analogous organoimido complexes containing nonoxidizable ligands. Organoimidorhenium-(V) dithiocarbamate complexes are well-known and can be prepared directly from the phosphine complexes. 9b,13,18 The replacement of chloride ligands is an additional important structural difference in these types of complexes. Solutions of complexes 3 and 4 in CH2Cl2 and CDCl₃ were monitored by UV-vis and ¹H NMR, respectively, and found to be stable in the presence of excess N-hydroxysuccinimide for at least 3 days at ambient temperature. The dimerization of the chloro complex 3 in the presence of H2O was described previously, and reactions of 3 with excess isopropylamine in CH2Cl2 containing trace amounts of water resulted in the formation of the bridging μ -oxo dimer O[ReO(S₂CNEt₂)₂]₂ and the hydrolyzed aniline derivative H₂NC₆H₄CO-NHCH(CH₃)₂. The dimeric succinimide ester complex 4 reacted with isopropylamine in CH₂Cl₂ with traces of H₂O to give only low yields of the desired amide product and was also accompanied by larger amounts of the hydrolysis product O[ReO(S₂CNEt₂)₂]₂. When the reaction of 4 with isopropylamine was performed in 10% DMF/H₂O, none of the desired bis-organoimido μ -oxo dimer $O[Re=N-C_6H_4CONHCH(CH_3)_2(S_2CNEt_2)_2]_2$ was isolated; only the hydrolyzed aniline and μ -oxo dimer O[ReO(S₂CNEt₂)₂]₂ were obtained. These results contrast with the general stability of complex 4 in aqueous DMF alone, but may simply reflect the enhanced nuleophilicity of water in the presence of an alkylamine base. Thus, while the organoimido dithiocarbamate complexes were unaffected by N-hydroxysuccinimide, these complexes were subject to the more severe problem of hydrolysis of the imido bond in the presence of amines.

Conclusions

A series of organoimido complexes possessing remotely functionalized succinimidyl ester groups were synthesized. The organoimidorhenium(V) N-hydroxy-succinimidyl ester phosphine complex 2 is an effective labeling agent for alkylamines when the reactions are carried out in aqueous DMF solution. A series of primary and secondary amines, amino acids, and a biotin—ethylenediamine derivative were successfully coupled with 2 to give the corresponding amides in good yields.

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These results demonstrate the stability of the organoimido linkage against competing hydrolysis to the oxo and in the presence of more basic alkylamines. When CH₂Cl₂ solvent was used, the amide formation was slower, and oxidation of the PPh3 ligands by the released N-hydroxysuccinimide caused decompostion of the organoimido complexes. The dithiocarbamate complexes 3 and 4 were unaffected by N-hydroxysuccinimide, but the organoimido groups were not stable under the reaction conditions with alkylamines and hydrolysis to the oxo complexes was observed. The problem of PPh₃ oxidation of 2 could potentially be avoided by preparing other activated carboxylic acid derivatives such as the pentachlorophenol or nitrophenol esters. The stability of the organoimido bond is dramatically affected by the nature of the ancillary ligands, and further efforts will attempt to develop systems with enhanced stability and water solubility.

Experimental Section

General Procedures. Reagents were purchased from Aldrich or Acros and used as received. Trichlorooxobis(triphenylphosphine)rhenium(V) was prepared according to the literature procedure. ¹⁹ ¹H NMR and ¹³C NMR spectra were recorded at 200 and 50 MHz, respectively, using CDCl₃ as solvent and internally referenced to TMS. ³¹P{¹H} NMR spectra were obtained at 161.9 MHz, referenced to an internal capillary containing 85% H₃PO_{4(aq)} (δ = 0). IR spectra were recorded as KBr pellets using a Perkin-Elmer 1720 X FT-IR spectrometer. UV—vis spectra were recorded using a Hewlett-Packard 8452A diode array spectrophotometer. Elemental analyses were performed by Desert Analytics, Tucson, AZ.

Synthesis of Cl₃(PPh₃)₂Re=N-C₆H₄CO₂N(COCH₂)₂ (2). Triphenylphosphine (1.57 g, 6 mmol) was added to a solution of N₃C₆H₄CO₂N(COCH₂)₂ (780 mg, 3.0 mmol) in benzene (200 mL) and stirred for 20 min at 25 °C, and trichlorooxobis-(triphenylphosphine)rhenium(V) (2.50 g, 3.0 mmol) was then added and the mixture heated to reflux for 1 h. The volatiles were removed in vacuo, and the product was recrystallized from dichloromethane by the addition of hexanes, filtered, and washed with Et₂O and hexanes to give a green solid, 2 (2.865 g, 91% yield): UV-vis (CH₂Cl₂) $\lambda = 234$, 266, 298 ($\epsilon = 16400$), 336 nm ($\epsilon = 14\,030$); FT-IR 1770, 1743, 1197, 745, 694, 521 cm^{-1} ; ¹H NMR δ 7.88-7.72 (m, 12H), 7.52 (d, J = 8.6 Hz, 2H), 7.35-7.20 (m, 18H), 6.88 (d, J=8.6 Hz, 2H), 2.91 (s, 4H); 13 C NMR δ 169.36, 160.49, 135.21, 132.22, 131.73, 131.17, 130.78, 128.22, 123.20, 121.31, 26.07; ${}^{31}P{}^{1}H$ NMR δ -23.66 (s). Anal. Calcd for C₄₇H₃₈Cl₃N₂O₄P₂Re: C, 53.80; H, 3.65; N, 2.67. Found: C, 53.73; H, 3.65 N, 2.66.

Synthesis of Cl[(Et)₂NCS₂]₂Re=N-C₆H₄CO₂N(COCH₂)₂ (3). A mixture of 2 (300 mg, 0.28 mmol) and tetraethylthiuramdisulfide (170 mg, 0.57 mmol) was heated at reflux in dry acetone (20 mL) under argon for 1 h, during which time the solution became dark green. The volume was reduced to 5 mL, and the flask was cooled in an ice bath to crystallize the complex. The complex was filtered and washed with cold acetone and Et₂O to give 3 (150 mg, 84% yield): UV-vis (CH₂-Cl₂): λ = 234, 272, 296 (ϵ = 24 045), 418 nm (ϵ = 4710); FT-IR 1769, 1741, 1531, 1073, 997 cm⁻¹; ¹H NMR δ 7.99 (d, J = 8.6 Hz, 2H), 7.54 (d, J = 8.6 Hz, 2H) 3.95-3.65 (m, 8H), 2.89 (s, 4H), 1.38 (t, J = 7.1 Hz, 12H); ¹³C NMR δ 239.88, 168.95, 161.12, 131.51, 123.81, 122.79, 45.16, 25.50, 12.42. Anal. Calcd for C₂₁H₂₈ClN₄O₄S₄Re: C, 33.61; H, 3.76; N, 7.47. Found: C, 33.37; H, 3.74; N, 7.41.

Synthesis of $O[((Et)_2NCS_2-)_2Re=N-C_6H_4CO_2N(CO-CH_2)_2]_2$ (4). To a solution of complex 3 (100 mg, 0.13 mmol) in

0.1% v/v aqueous acetone (20 mL) was added sodium carbonate (400 mg), and the mixture was heated at reflux for 2 h. The reaction mixture was filtered and concentrated in vacuo, and the crude product was isolated by addition of hexanes to a dichloromethane solution and purified by recrystallization from hexanes/CH₂Cl₂ to give the product 4 (70 mg, 73% yield): UV-vis (CH₂Cl₂) $\lambda = 244$, 268, 348, 444 nm; FT-IR 1780, 1744, 1500, 1072, 684 cm⁻¹; ¹H NMR δ 7.88 (d, J = 8.7 Hz, 4H), 7.28 (d, J = 8.2 Hz, 4H), 3.95-3.55 (m, 16H), 2.87 (s, 8H), 1.37 (t, J = 7.0 Hz, 24H); ¹³C NMR δ 239.47, 169.27, 164.24, 130.99, 124.55, 118.70, 43.70, 25.53, 12.63.

Synthesis of Cl₃(Ph₃P)₂Re=N-C₆H₄CONHCH(CH₃)₂ (5a). Isopropylamine (50 μ L, 1 mmol) was added to a solution of 2 (105 mg, 0.1 mmol) in DMF (4 mL) and stirred at 25 °C for 10 min. Water (36 mL) was added while stirring vigorously to precipitate the complex. The product was filtered, washed with water and hexanes, and then recrystallized from dichloromethane by adding hexanes to give the green solid **5a** (70 mg, 70% yield): UV-vis (CH₂Cl₂) λ = 234, 264, 346 nm (ϵ = 13000); FT-IR 1639, 1525, 1435, 1093, 745, 694, 521 cm⁻¹; ¹H NMR δ 7.85-7.70 (m, 12H), 7.38-7.15 (m, 20H), 6.88 (d, J = 8.4 Hz, 2H), 5.76 (m, 1H), 4.23 (m, 1H), 1.26 (d, J = 6.4 Hz, 6H); ¹³C NMR δ 163.62, 158.19, 135.33, 133.29, 132.17, 131.67, 130.70, 128.22, 121.76, 42.81, 23.13; ³¹P{¹H} NMR δ -21.05 (s). Anal. Calcd for C₄₆H₄₂Cl₃N₂OP₂Re·0.5CH₂Cl₂: C, 53.86; H, 4.15; N, 2.70. Found: C, 53.89; H, 4.09; N, 2.77.

Synthesis of Cl₃(Ph₃P)₂Re=N-C₆H₄CONHCH(CH₃)CH-(CH₃)₂ (5b). The procedure described for the synthesis of 5a was repeated using 2 (105 mg, 0.10 mmol) and 2-amino-3-methylbutane (87 mg, 1.00 mmol) to give the green solid 5b (70 mg, 69% yield): UV-vis (CH₂Cl₂) λ = 234, 264, 346 nm (ϵ = 11 437); FT-IR 1655, 1536, 1435, 1093, 745, 694, 521 cm⁻¹; ¹H NMR δ 7.88-7.68 (m, 12H), 7.40-7.10 (m, 20H), 6.88 (d, J = 7.4 Hz, 2H), 5.75 (m, 1H), 4.02 (m, 1H), 1.80 (m, 1H), 1.17 (d, J = 6.6 Hz, 3H), 0.95 (d, J = 6.0 Hz, 6H); ¹³C NMR δ 165.22, 157.61, 134.73, 131.51, 131.09, 130.09, 127.59, 127.11, 121.20, 50.71, 32.86, 18.51, 17.36; ³¹P{¹H} NMR δ -19.46.

Synthesis of Cl₃(Ph₃P)₂Re=N-C₆H₄CONH(CH₂)₂Ph (5c). The procedure described for the synthesis of **5a** was repeated using **2** (50 mg, 0.05 mmol) and phenethylamine (50 μ L, 0.50 mmol) to give the green complex **5c** (35 mg, 67%): UV-vis (CH₂Cl₂) λ = 232, 264, 344 nm (ϵ = 10 598); FT-IR 1665, 1541, 1093, 747, 694, 521 cm⁻¹; ¹H NMR δ 7.90-7.70 (m, 12H), 7.45-7.15 (m, 23H), 7.08 (d, J = 8.4 Hz, 2H), 6.82 (d, J = 8.4 Hz, 2H), 5.96 (m, 1H), 3.67 (q, J = 6.7 Hz, 2H), 2.92 (t, J = 6.7 Hz, 2H); ¹³C NMR δ 166.48, 158.10, 139.12, 135.30, 132.08, 131.69, 131.15, 130.65, 129.23, 128.21, 127.87, 127.19, 121.77, 41.75, 35.87; ³¹P{¹H} NMR δ -19.83 (s).

Synthesis of Cl₃(Ph₃P)₂Re=N-C₆H₄CONHCH₂Ph (5d). The procedure described for the synthesis of **5a** was repeated using **2** (105 mg, 0.1 mmol) and benzylamine (110 μ L, 1 mmol), and the crude product was recrystallized by slow diffusion of ethanol into a dichloromethane solution to give the green complex **5d** (75 mg, 72% yield): UV-vis (CH₂Cl₂) λ = 234, 264, 344 nm (ϵ = 13 398); FT-IR 1656, 1093, 745, 694, 521 cm⁻¹; ¹H NMR δ 7.82-7.73 (m, 12H), 7.40-7.30 (m, 5H), 7.26-7.17 (m, 20H), 6.86 (d, J = 8.4 Hz, 2H), 6.35 (t, J = 5.2 Hz, 1H), 4.58 (d, J = 5.2 Hz, 2H); ¹³C NMR δ 166.30, 158.27, 138.19, 135.20, 133.97, 132.66, 132.06, 131.64, 130.63, 129.31, 128.56, 120.17, 121.75, 44.71; ³¹P{¹H} NMR δ -21.29 (s). Anal. Calcd for C₅₀H₄₂Cl₃N₂O₂P₂Re-0.25CH₂Cl₂: C, 56.74; H, 3.98. Found: C, 56.92; H, 3.60.

Synthesis of $\text{Cl}_3(\text{Ph}_3\text{P})_2\text{Re}=\text{N-C}_6\text{H}_4\text{CON}(\text{CH}_2)_4$ (5e). The procedure described for the synthesis of 5a was repeated using 2 (50 mg, 0.05 mmol) and pyrrolidine (50 μL , 0.50 mmol), and the crude product was recrystallized by slow diffusion of ethanol into a dichloromethane solution to give the green complex 5e (35 mg, 70% yield) and recrystallized from dichloromethane and ethanol: UV-vis (CH₂Cl₂) λ = 232, 264, 344 nm (ϵ = 10 486); FT-IR 1628, 1092, 745, 694, 521 cm⁻¹; ¹H NMR δ 7.85-7.65 (m, 12H), 7.30-7.15 (m, 18H), 6.93 (d, J =

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8.0 Hz, 2H), 6.86 (d, J=8.0 Hz, 2H), 3.58 (t, J=6.4 Hz, 2H), 3.22 (t, J=6.4 Hz, 2H), 2.05–1.82 (m, 4H); 13 C NMR δ 168.65, 156.85, 135.31, 132.16, 131.66, 131.20, 130.60, 128.14, 121.64, 49.82, 46.73, 26.81, 24.79; 31 P{ 1 H} NMR δ –21.00 (s). Anal. Calcd for C₄₇H₄₂Cl₃N₂OP₂Re·0.5CH₂Cl₂: C, 54.39; H, 4.10; N, 2.67. Found: C, 54.55; H, 4.13; N, 2.66.

Synthesis of Cl₃(Ph₃P)₂Re=N-C₆H₄CON(CH₂CH₂)₂O (5f). The procedure described for the synthesis of **5a** was repeated using **2** (105 mg, 0.10 mmol) and morpholine (88 μ L, 1.00 mmol), and the precipitated product was recrystallized by slow diffusion of ethanol into a dichloromethane solution to give the green complex **5f** (80 mg, 78%): UV-vis (CH₂Cl₂) λ = 234, 264, 344 nm (ϵ = 13 922); FT-IR 1636, 1093, 746, 694, 521 cm⁻¹; ¹H NMR δ 7.85-7.65 (m, 12H), 7.30-7.15 (m, 18H), 6.88 (d, J = 8.0 Hz, 2H), 6.81 (d, J = 8.0 Hz, 2H), 3.99 (m, 2H), 3.80-3.50 (m, 4H), 3.25 (m, 2H); ¹³C NMR δ 169.39, 156.93, 135.33, 132.22, 131.73, 131.17, 130.62, 128.16, 121.85, 61.12, 48.64; ³¹P{¹H} NMR δ -21.36 (s). Anal. Calcd for C₄₇H₄₂-Cl₃N₂O₂P₂Re-0.5CH₂Cl₂: C, 53.57; H, 4.04; N, 2.63. Found: C, 53.16; H, 3.92; N, 2.64.

Synthesis of Cl₃(Ph₃P)₂Re=N-C₆H₄CONHCH(CH₃)CO₂Et (5g). Triethylamine (300 μ L, 2.00 mmol) was added to a solution of alanine ethyl ester hydrochloride (303 mg, 2.00 mmol) in DMF (4 mL), and the solution was stirred for 15 min. A solution of complex 2 (210 mg, 0.20 mmol) in DMF (4 mL) was then added, and the mixture was stirred for 10 min. Water (72 mL) was added, and the precipitated product 5g was isolated by filtration and was subsequently recrystallized by addition of hexanes to a CH₂Cl₂ solution (140 mg, 67% yield): UV-vis (CH₂Cl₂) $\lambda = 232$, 264, 342 nm ($\epsilon = 12\,000$); FT-IR 1737, 1666, 1092, 750, 693, 521 cm⁻¹; 1 H NMR δ 7.85-7.68 (m, 12H), 7.35-7.15 (m, 20H), 6.87 (d, J = 8.4 Hz, 2H), 6.84(d, J = 7.2 Hz, 1H), 4.70 (m, 1H), 4.25 (q, J = 7.0 Hz, 2H), 1.51 (d, J = 6.8 Hz, 3H), 1.32 (t, J = 7.0 Hz, 3H); ¹³C NMR δ 172.90, 165.28, 157.90, 134.70, 131.54, 131.11, 130.70, 130.09, 127.59, 121.19, 61.77, 48.63, 18.38, 14.01; ${}^{31}P{}^{1}H{}^{1}NMR$ δ -21.50 (s). Anal. Calcd for C₄₈H₄₄Cl₃N₂OP₂Re: C, 54.83; H, 4.22; N, 2.66. Found: C, 54.83; H, 4.18; N, 2.79.

Synthesis of Cl₃(Ph₃P)₂Re=N-C₆H₄CONHCH(CH₂SH)-CO₂Et (5h). The procedure described for the synthesis of 5g was repeated using the cysteine ethyl ester hydrochloride (370 mg, 2.00 mmol), DMF (3 mL), triethylamine (300 μ L, 2.00 mmol), 2 (210 mg, 0.20 mmol), DMF (3 mL), and water (54 mL). The product was recrystallized from dichloromethane and hexanes to give the green complex 5h (160 mg, 74% yield): UV-vis (CH₂Cl₂) λ = 234, 264, 336 nm (ϵ = 14 870); FT-IR 1737, 1665, 1093, 746, 694, 521 cm⁻¹; ¹H NMR δ 7.82-7.70 (m, 12H), 7.35-7.15 (m, 20H), 6.88 (d, J = 8.0 Hz, 2H), 4.99 (m, 1H), 4.35-4.25 (m, 3H), 3.20-3.05 (m, 2H), 1.34 (t, J = 7.6 Hz, 3H); ³¹P{¹H} NMR δ -21.23 (s).

Synthesis of $Cl_3(Ph_3P)_2Re=N-C_6H_4CONH(CH_2)_2NH-Biotin (5i)$. The procedure described for the synthesis of 5g was repeated with 6 (108 mg, 0.38 mmol), DMF (6 mL), triethylamine (86 μ L, 0.57 mmol), complex 2 (200 mg, 0.19 mmol), DMF (6 mL), and water (108 mL). The product was recrystallized from dichloromethane and hexanes to give the

green complex 5i (107 mg, 49%): UV-vis (CH₂Cl₂) λ $\stackrel{c}{=}$ 23 $\stackrel{c}{\times}$, 264, 342 nm (ϵ = 7045); FT-IR 1705, 1655, 1544, 1435, 1093, 747, 694, 521 cm⁻¹; 1 H NMR (CDCl₃+CD₃OD) δ 7.88-7.68 (m, 12H), 7.48-7.22 (m, 20H), 6.88 (d, J = 8.0 Hz, 2H), 4.55-4.40 (m, 1H), 4.35-4.22 (m, 1H), 3.58-3.32 (m, 4H), 3.20-2.65 (m, 3H), 2.23 (t, J = 8 Hz, 2H), 1.85-1.25 (m, 4H); 13 C NMR (CDCl₃+CD₃OD) δ 175.71, 167.42, 157.95, 135.06, 133.79, 131.87, 131.42, 130.57, 128.66, 128.07, 121.67, 62.25, 60.50, 55.73, 40.99, 40.58, 39.21, 35.87, 28.33, 25.66; 31 P{ 1 H} NMR δ -20.85 (s).

X-ray Structure Determination of Cl₃(PPh₃)₂Re=N- $C_6H_4CO_2N(COCH_2)_2 \cdot (CH_2Cl_2)_{1.43}(EtOH)_{0.57}$. Complex 2 was crystallized by slow diffusion of ethanol into a CH2Cl2 solution of 2 at room temperature to give green, rod-shaped crystals of the complex 2·(CH₂Cl₂)_{1.43}·(EtOH)_{0.57}. A suitable crystal was suspended in viscous mineral oil, mounted on a glass fiber, and cooled to -70 °C. Data collection was performed by a Siemens R3m/V diffractometer operating in the θ -2 θ scan mode with graphite-monochromated Mo K α radiation (λ = 0.710 73 Å) as previously described.20 Intensities were corrected for Lorentz and polarization effects, and empirical absorption corrections were applied based on a set of $\boldsymbol{\psi}$ scans. The structure was solved by direct methods using Siemens' SHEXTL PLUS structure package. The unit cell and lack of systematic absences indicated a Laue symmetry of triclinic and the space group P1. Two solvent molecules are included in the asymmetric unit, with one exhibiting complex disorder. This site refined adequately for 57% occupancy by EtOH and 43% occupancy by CH2Cl2, disordered over three positions. The structure was refined with full-matrix least-squares, and all non-hydrogen atoms in 2 and the ordered CH2Cl2 were refined anisotropically. The positions of the hydrogen atoms were calculated (C-H = 0.95 Å) and allowed to ride isotropically on their respective carbons atoms during refinement. Refinement of the data converged with a goodness-of-fit of 1.20 and final residuals (for 7463 data with $F \ge 4\sigma_F$) of R = 3.44% and $R_{\rm w} = 4.19\%$. Crystallographic data are presented in Table 1; selected bond distances and angles are given in Table 2.

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Supporting Information Available: Complete tabulations of crystallographic data, bond lengths and angles, atomic coordinates, and thermal parameters, a completely labeled ball-and-stick diagram, and copies of ¹H, ¹³C{¹H}, and ³¹P{¹H} NMR spectra for complexes **4**, **5b**, **5c**, **5h**, and **5i**. This material is available free of charge via the Internet at http://pubs.acs.org.

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Trichlorooxo(triphenylphosphine)(triphenylphosphine oxide)rhenium(V)

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Abstract

The title complex, $[ReCl_3O(C_{18}H_{15}OP)(C_{18}H_{15}P)]$, is produced in a reaction between $[Re(O)Cl_3(PPh_3)_2]$ and ethyl 2-hydroxymethyl sulfoxide. The structure is compared to that of $[Re(O)Cl_3(PPhEt_2)(OPPhEt_2)]$. The Re—Cl distances are shorter $[2.361\ (2)-2.384\ (2)\ Å]$ and the Re—P distance is longer $[2.506\ (2)\ Å]$ in the title complex.

Comment

A variety of interesting and synthetically useful O-atom transfer reactions from sulfoxide substrates catalyzed by the precursor complex [Re(O)Cl₃(PPh₃)₂] have recently been reported (Bryan *et al.*, 1987; Arterburn &

Perry, 1996; Arterburn & Nelson, 1996; Arterburn et al., 1997). This compound was known to react with dimethyl sulfoxide in the presence of hydrochloric acid to form the mixed dimethylsulfide–triphenylphosphine oxide complex [Re(O)Cl₃(SMe₂)(OPPh₃)]. However, no intermediate complexes from the catalytic reactions in organic solvents have yet been structurally identified. The precursor complex was found to react with one equivalent of ethyl 2-hydroxyethyl sulfoxide, CH₃CH₂S(O)CH₂CH₂OH, at ambient temperature to give a purple solution. The title complex, (I), and amorphous purple solids were obtained following precipitation with diethyl ether. These results are consistent with catalytic pathways involving coordinated Re^V–oxo complexes as intermediates.

A distorted octahedral coordination geometry is observed around Re (Fig. 1). The major distortion is an increase in the O1—Re—Cl angles, which is commonly observed in octahedral complexes containing a

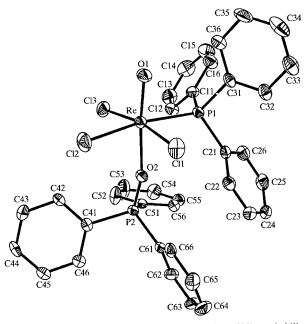


Fig. 1. The molecular structure of (I) showing 50% probability displacement ellipsoids. H atoms have been omitted for clarity.

4 A A

multiply bonding ligand (Nugent & Mayer, 1988; Bryan et al., 1987). The location of softer ligands cis to the oxo ligand is also often observed (Bryan et al., 1987). The short Re-oxo (O1) bond length of 1.669 (4) Å is consistent with a strong multiple bond (Nugent & Mayer, 1988). The structure of (I) is very similar to the closely related [Re(O)Cl₃(PPhEt₂)(OPPhEt₂)] (Sergienko, 1994; Sergienko et al., 1982). The Re—Cl distances are shorter [2.361 (2)-2.384 (2) Å] in (I) than in [Re(O)Cl₃(PPhEt₂)(OPPhEt₂)] [2.394 (4)-2.407 (4) Å], while the Re—P distance is longer [2.506 (2) versus 2.464 (4) Å]. These differences can be understood by the fact that PPh₃ is a weaker donor ligand than PPhEt₂.

Experimental

The preparation of (I) has been previously reported (Grove & Wilkinson, 1966; Bryan *et al.*, 1987). Green crystals suitable for X-ray diffraction were prepared by slow diffusion of methyl *tert*-butyl ether into an acetone solution of the complex.

Crystal data

Data collection

Nonius CAD-4 diffractometer ω scans Absorption correction: ψ scan (Siemens, 1997) $T_{\min} = 0.274, T_{\max} = 0.393$ 8509 measured reflections 7561 independent reflections 5690 reflections with	$R_{\rm int} = 0.039$ $\theta_{\rm max} = 27.5^{\circ}$ $h = 0 \rightarrow 24$ $k = -12 \rightarrow 5$ $l = -25 \rightarrow 23$ 3 standard reflections frequency: 120 min intensity decay: 3%
$I > 2\sigma(I)$	

Refinement

+ 4.6318P] where $P = (F_o^2 + 2F_c^2)/3$

<i>j</i>	
Refinement on F^2	$(\Delta/\sigma)_{\text{max}} = -0.001$ $\Delta\rho_{\text{max}} = 2.40 \text{ e Å}^{-3}$
R(F) = 0.040	$\Delta \rho_{\text{max}} = 2.40 \text{ e Å}^{-3}$
$wR(F^2) = 0.099$	$\Delta \rho_{\min} = -1.47 \text{ e Å}^{-3}$
S = 1.074	Extinction correction: none
7561 reflections	Scattering factors from
397 parameters	International Tables for
H atoms not refined	Crystallography (Vol. C)
$w = 1/[\sigma^2(F_o^2) + (0.0502P)^2]$	

Table 1. Selected geometric parameters (Å, °)

Re-CI1	2.373 (2)	Rc—Ol	1.669 (4)
ReC12	2.361(2)	RcO2	2.082(4)
Re-C13	2.384(2)	P2O2	1.508 (4)
Re-P1	2.506(2)		
CI1—Re—CI2	86.53 (6)	Cl2—Re—O2	87.33 (11)
CI1—Re—CI3	169.74 (6)	Cl3—Rc—Pl	95.09 (5)
Cl1RePl	89.69 (5)	Cl3—Re—O1	94.07 (19)
Cl1—Re—O1	95.25 (19)	Cl3—Re—O2	84.17 (12)
Cl1—Re—O2	87.23 (12)	P1—Re—O1	86.82 (16)
C12-ReC13	87.50(6)	P1ReO2	84.90 (11)
C12-Re-P1	171.52 (6)	O1—Re—O2	171.35 (18)
C12-Re-O1	101.07 (16)	Re—O2—P2	165.1 (3)

Anisotropic displacement parameters were used for all non-H atoms. All H atoms were placed in calculated positions (C—H = 0.95 Å), refined using a riding model, and given isotropic displacement parameters equal to 1.2 times the equivalent isotropic displacement parameter of the atom to which they are attached. The highest eight peaks in the final difference map (electron density greater than $0.8 \, \text{e} \, \text{Å}^{-3}$) are located approximately $1 \, \text{Å}$ from Re.

Data collection: *CAD-4-PC* (Enraf–Nonius, 1993). Cell refinement: *CAD-4-PC*. Data reduction: *XCAD4* (Harms, 1995). Program(s) used to solve structure: *SHELXS*86 (Sheldrick, 1990). Program(s) used to refine structure: *SHELXL93* (Sheldrick, 1993). Molecular graphics: *SHELXTL* (Siemens, 1997). Software used to prepare material for publication: *PLA-TON* (Spek, 1998).

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